MicroRNAs Are Absorbed in Biologically Meaningful Amounts from Nutritionally Relevant Doses of Cow Milk and Affect Gene Expression in Peripheral Blood Mononuclear Cells, HEK-293 Kidney Cell Cultures, and Mouse Livers

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Abstract

Background: MicroRNAs (miRNAs) regulate genes in animals and plants and can be synthesized endogenously. In milk, miRNAs are encapsulated in exosomes, thereby conferring protection against degradation and facilitating uptake by endocytosis. The majority of bovine miRNAs have nucleotide sequences complementary to human gene transcripts, suggesting that miRNAs in milk might regulate human genes.

Objectives: We tested the hypotheses that humans absorb biologically meaningful amounts of miRNAs from nutritionally relevant doses of milk, milk-borne miRNAs regulate human gene expression, and mammals cannot compensate for dietary miRNA depletion by endogenous miRNA synthesis.

Methods: Healthy adults (3 men, 2 women; aged 26–49 y) consumed 0.25, 0.5, and 1.0 L of milk in a randomized crossover design. Gene expression studies and milk miRNA depletion studies were conducted in human cell cultures and mice, respectively. For comparison, feeding studies with plant miRNAs from broccoli were conducted in humans.

Results: Postprandial concentration time curves suggest that meaningful amounts of miRNA (miR)-29b and miR-200c were absorbed; plasma concentrations of miR-1 did not change (negative control). The expression of runt-related transcription factor 2 (RUNX2), a known target of miR-29b, increased by 31% in blood mononuclear cells after milk consumption compared with baseline. When milk exosomes were added to cell culture media, mimicking postprandial concentrations of miR-29b and miR-200c, reporter gene activities significantly decreased by 44% and 17%, respectively, compared with vehicle controls in human embryonic kidney 293 cells. When C57BL/6J mice were fed a milk miRNA-depleted diet for 4 wk, plasma miR-29b concentrations were significantly decreased by 61% compared with miRNA-sufficient controls, i.e., endogenous synthesis did not compensate for dietary depletion. Broccoli sprout feeding studies were conducted as a control and elicited no detectable increase in Brassica-specific miRNAs.

Conclusion: We conclude that miRNAs in milk are bioactive food compounds that regulate human genes. J. Nutr. 144:1495–1500, 2014.

Introduction

MicroRNAs (miRNAs) are small, noncoding RNAs (~22 nucleotides in mature miRNAs) that may silence genes via destabilizing complementary mRNA sequences or preventing translation of mRNAs (1,2). The nucleotide sequence in the seed region in miRNAs (nucleotides 2–7) is of particular importance for binding to target mRNA (3); imperfect pairing of sequences in the seed region in miRNA to mRNA impairs gene down-regulation at the protein or RNA level (4). miRNAs are encoded by their own genes, introns, or exons of long nonprotein-coding transcripts (5). Traditionally, miRNAs are considered endogenous regulators of genes, i.e., miRNAs synthesized by human cells regulate the expression of genes in that host. We challenged this paradigm and tested the hypothesis that humans absorb miRNAs from milk in meaningful quantities and milk-borne...
miRNAs elicit biologically meaningful changes in human gene expression.

There is precedent for the bioavailability of dietary miRNAs in humans. A recent report suggested that miRNA (miR)-168a from rice (Oryza sativa; osa-miR-168a) is detectable in human and animal sera, and osa-miR-168a decreases the expression of LDL receptor adapter protein 1 mRNA, thereby inhibiting LDL receptor expression in mouse liver (6). Note that the serum concentration of osa-miR-168a is only ~3 fmol/L in humans and that the bioavailability of plant-borne miRNAs in humans is controversial (6–10).

This study focused on milk as a source of dietary miRNAs on the basis of the rationale that a large fraction of miRNAs in milk is contained in exosomes, conferring protection against degradation (11,12), Americans consume 88 L of milk annually (13), and cow milk contains large quantities of 245 miRNAs (14,15). This study of milk-borne miRNAs was modeled primarily on miR-29b on the basis of the rationale that the nucleotide sequence of bovine miR-29b is identical to that of human miR-29b (16), and miR-29b increases bone mineralization in humans through promoting a gain in osteoblast differentiation (17) and a loss in osteoclast differentiation and function (18). In select experiments, miR-200c was included because its concentration is 8 times that of miR-29b in cow milk (15). The nucleotide sequence of bovine miR-200c is identical to that of human miR-200c (16), and miR-200c decreases cancer risk by targeting the transcription factor zinc finger E-box binding homeobox 1 (ZEB1); miR-200c-dependent loss of ZEB1 induces E-cadherin expression, thereby limiting epithelial-to-mesenchymal transition, a key event in metastasis (19,20).

This study was guided by the following aims. First, we determined whether humans absorb quantitatively meaningful amounts of miRNAs from nutritionally relevant doses of milk and characterized the bioavailability of milk miRNAs by using pharmacokinetics protocols. Second, we assessed the effects of physiologically relevant miRNA concentrations on the expression of endogenous genes in human peripheral blood mononuclear cells (PBMCs) and reporter genes in human cell cultures. Third, we determined whether mammals compensate for dietary miRNA deficiency by an increased synthesis of endogenous miRNAs in a mouse miRNA depletion study. Fourth, we conducted a broccoli feeding study to determine whether plant-borne miRNAs are bioavailable in humans.

**Materials and Methods**

**Human feeding study.** Five apparently healthy adults (3 men, 2 women) participated in a milk feeding study that used 3 doses of milk in a randomized crossover design with a washout period of at least 1 wk between doses. Exclusion criteria included pregnancy, smoking, milk allergies, and self-reported health problems. Preliminary studies suggested that plasma miRNA concentrations remain higher than baseline concentrations until 9 h after milk consumption; participants were instructed not to consume milk and dairy products for 12 h before the milk meal and during the period in which blood samples were collected. Doses of milk were normalized by total body water of participants, which is the suspected volume of distribution for miRNAs. The estimation of total body water was calculated as previously described (21). Normalization of doses by body water resulted in doses (means ± SEMs) of 0.218 ± 0.018, 0.436 ± 0.037, and 0.872 ± 0.073 L, representing the equivalent of 0.25, 0.5, and 1.0 L in a 26- y old reference male (75 kg weight, 1.83 m height). Twenty milliliters of blood was collected before milk consumption (baseline; time = 0 h) and at timed intervals (1–3, 6, 9, and 24 h) after the milk meal. PBMCs and plasma were collected by using gradient centrifugation as described previously (22) and frozen at −80°C until analysis. This protocol was approved by the University of Nebraska–Lincoln Institutional Review Board, and all participants provided signed informed consent forms before participation.

**Real-time qPCR.** The sequences of mature bovine miR-29b (bta-miR-29b), miR-200c (bta-miR-200c), and miR-1 (bta-miR-1; control) are identical to their human and murine orthologs, and therefore real-time qPCR amplification targeted the mature human miR-29b (hsa-miR-29b-3p), miR-200c (hsa-miR-200c-3p), and miR-1 (hsa-miR-1-3p) that originate in the 3′p arm of pre-miRNA (23). RNA was isolated from human and murine (see below) plasma by using the NucleoSpin miRNA plasma kit (Machery-Nagel). Reverse transcription was performed by using the miScript II RT kit (Qiagen). Real-time qPCR was performed by using miScript SYBR Green (Qiagen) and the universal reverse primer included in the kit plus primers specific for individual miRNA (Supplemental Table 1). miR-1 is not detectable in milk (data not shown) and served as a negative control. Five attomoles of synthetic miRNA, miR-29b (5×10⁻⁸ M), and miR-1 (5×10⁻⁸ M), was added to each sample after denaturation of plasma with lysin buffer and served as an external standard for calibration of real-time qPCR.

**Pharmacokinetics analysis.** AUCs for plasma miRNAs were calculated by using the linear trapezoidal rule to assess the apparent bioavailability of miRNAs (24). Plasma concentrations of miR-29b and miR-200c returned to baseline values 9 h after consumption of the 2 lowest doses of milk. For the calculation of AUC, baseline values were subtracted from postprandial concentrations and the plasma concentrations from the first 9 h after milk meals were used. The maximal plasma concentration (Cmax) and the time of peak concentration (tmax) were obtained by visual inspection of the plasma time curves measured for miR-29b and miR-200c. Cmax of an orally administered compound is a marker of bioavailability and rate of absorption, whereas tmax is a marker of the approximate absorption site (25).

**Gene expression.** The effects of milk-borne miRNAs on the expression of human genes were assessed in cell cultures and PBMCs from the human milk feeding studies. For studies in cell cultures, human embryonic kidney (HEK)-293 cells (American Type Culture Collection) were cultured in MEM containing 10% exosome-depleted FBS, 0.1% sodium pyruvate, 0.1%, and 100 mg/L streptomycin for 10 d. Bovine serum was depleted of exosomes by ultracentrifugation at 130,000 × g for 4 h, leading to the removal of 97% and 91% of miR-29b and miR-200c, respectively. Cells were transfected with miRNA reporter genes as described previously (26). Forty-eight hours after transfection, exosome-depleted media were replaced with exosome-sufficient media at final concentrations of 600 fmol/mL miR-29b or 1000 fmol/mL miR-200c. Exosome-sufficient media were prepared by using exosomes collected from cow milk as previously described (27). The concentrations of miR-29b and miR-200c in exosomes were quantified by using real-time qPCR, and exosomes were added back to culture media to produce the desired concentrations of miRNAs. Reporter genes for miR-29b and miR-200c were created by inserting the 3′-untranslated regions (3′-UTRs) from genes collagen, type I, α1 (COL1A1; 3 miR-29b binding sites), and ZEB1 (2 miR-200c binding sites, respectively), downstream of the luciferase (LUC) reading frame, driven by a cytomegalovirus promoter, thereby creating plasmids LUC-mir-29b and LUC-mir-200c (Supplemental Fig. 1). LUC-mir-200c was obtained from Dr. Thomas Grabetz (University of Freiburg, Germany) and is denoted as ZEB1 3′UTR-Luc in the original publication (20). LUC-mir-29b was created by digesting LUC-mir-200c with MluI and Hind III to remove the ZEB1 sequence. The 3′-UTRs of COL1A1 were amplified by PCR (Supplemental Table 1) by using IMR-90 human lung fibroblast DNA as a template and ligated into the reporter plasmid by using MluI and Hind III. Cells were cotransfected with plasmid Rous sarcoma virus-β-galactosidase and luciferase plasmids to assess transfection efficiency (28). Reporter gene activities were normalized by transfection efficiency. Our rationale for choosing HEK-293 cells was that these cells can be transfected with near 100% efficiency and express COL1A1 in meaningful quantities (29,30).

For studies in human PBMCs, a preliminary time-response screen was conducted to determine the time point when the effects of milk on the expression of runt-related transcription factor 2 (RUNX2) and ZEB1 miRNAs were maximal. RUNX2 is a downstream target of miR-29b-dependent signaling pathways; note that mir-29b is a positive
regulator of the expression of RUNX2 (17). Our preliminary assessment suggested that changes in mRNA abundance were maximal 6 h after milk consumption (Supplemental Fig. 2; positive and negative changes for RUNX2 and ZEB1 mRNAs, respectively). Therefore, subsequent assays of RUNX2 and ZEB1 mRNAs were conducted by using samples from the time point 6 h after the milk meal by using real-time qPCR (Supplemental Table 1) and the ΔΔG method (31); GAPDH was used to normalize for amplification efficiency (32). The abundance of miRNAs was measured as described for plasma samples, except that normalization was performed by using U6 rather than miSPIKE.

Mouse feeding study. An miRNA depletion study was conducted in mice to determine whether endogenous miRNA synthesis compensates for dietary deficiency. Ten female C57BL/6j mice (Jackson Laboratory; stock 000664), aged 3 wk, were randomly divided between a milk miRNA-depleted treatment group (denoted ExoMinus) and a milk miRNA-sufficient control group (ExoPlus). Diets were based on the AIN-93G formula with the following modifications and were fed for 4 wk (33). In the ExoMinus diet, exosome-depleted fat-free cow milk was substituted for casein and cornstarch in the AIN-93G diet to provide 10% of total calories (Supplemental Table 2); the remainder of casein in the AIN-93G diet was replaced with soy protein (Bob’s Red Mill) to eliminate any milk-borne miRNAs. Milk was depleted of exosomes by ultracentrifugation for 60 min (WVR Aquasonic 250T) and incubation at 37°C for 60 min before lyophilization. Ten percent of calories represent a consumption of ~0.5 L of fat-free milk by an adult on the basis of 352 kcal/L milk and an energy intake of 1760 kcal (34). The ExoPlus control diet was prepared by using exosome-containing milk powder but was otherwise identical to the ExoMinus diet. Components were blended and dried at 40°C overnight. No miR-29b was detectable in the ExoMinus diet, whereas the ExoPlus diet contained ~83 fmol/g miR-29b, assessed by real-time qPCR. All mouse protocols used in this study were approved by the University of Nebraska–Lincoln Institutional Animal Care and Use Committee. After 4 wk of feeding miRNA-defined diets, mice were killed by carbon dioxide, and blood was collected by depilation, and flash-frozen in liquid nitrogen. Ten milligrams of liver was homogenized by using lysis buffer in the NucleoSpin miRNA kit (Macherey-Nagel), and mRNA was purified following the manufacturer’s instructions. Reverse transcription was performed with the High Capacity RNA-to-cDNA kit (Life Technologies) for subsequent analysis of Kruppel-like factor 8 (KFL8) expression using real-time qPCR (Supplemental Table 1).

Broccoli-borne miRNAs in humans. The bioavailability of plant-borne miRNAs is controversial (6–10). We used plasma samples archived from a previous broccoli sprout feeding study in humans to determine whether broccoli-borne miRNAs are detectable in prandial plasma samples (35). In this previous study, 8 healthy adults were provided 34, 68, and 102 g of broccoli sprouts, and timed samples were collected at t = 0, 2, 4, 8, and 24 h. Here, we analyzed samples collected at t = 0 and 4 h from the highest dose of broccoli sprouts for miR-824 and miR-167a in 4 randomly selected participants. Total RNA was isolated from 75 mg of broccoli sprouts by using Trizol. Samples were analyzed for miR-824 and miR-167a by using real-time qPCR (Supplemental Table 1). As of today, there is no evidence that humans synthesize miR-824 and miR-167a (36), i.e., the Brassica-specific miR-824 and plant-specific miR-167a are good markers of broccoli-borne miRNAs in human blood.

Statistical analyses. Homogeneity of variances was tested by using Bartlett’s test. Variances were heterogeneous for miRNAs in mouse plasma, i.e., murine plasma data were log-transformed before subsequent statistical analysis. AUCs were calculated by using GraphPad Prism 6 (GraphPad Software). Pharmacokinetics data were analyzed by using repeated-measures ANOVA and Fisher’s protected least significant difference test for post hoc comparisons (37). PBMC gene expression data were analyzed by using the paired t test, whereas plasma miRNA concentrations and liver gene expression in mice were analyzed by using the Wilcoxon signed-rank test. StatView 5.0.1 (SAS Institute) was used for statistical analyses. Differences were considered significant if P < 0.05.

Results

Bioavailability of milk-borne miRNAs in humans. Cow milk (1% fat) contained 148 ± 42 and 680 ± 151 pmol/L of miR-29b and miR-200c, respectively. Humans absorbed considerable amounts of miR-29b from the cow milk (Fig. 1A). In contrast, the plasma concentration of miR-1 did not change after the consumption of 1 L of milk (negative control). Plasma miR-29b concentrations returned to baseline concentrations 9 h after the milk meals for the 0.25- and 0.5-L doses and 24 h after the 1-L dose. The AUC for miR-29b exhibited a linear dose-response relation regarding the amount of milk consumed (Table 1). Likewise, Cmax increased linearly with the amount of milk consumed, if Cmax was corrected for baseline concentrations of miR-29b. The tmax occurred 3.4 to 4.2 h after the milk meal. The miRNA miR-200c is another miRNA present in cow milk. Humans also absorbed considerable amounts of miR-200c from cow milk (Fig. 1B). Although the postprandial AUC for miR-200c was significantly higher for the 0.5- and 1.0-L doses of milk than for the 0.25-L dose, the AUC did not exhibit a linear dose-response relation; the AUCs were not significantly different between the 0.5- and the 1.0-L doses (Table 1).

Effects of milk miRNAs on the concentrations of miRNAs in human PBMCs. Milk feeding elicited an increase in the concentrations of miRNAs in human PBMCs that was significant only for miR-200c and trended toward significance for miR-29b (P = 0.09) (Fig. 2).

Effects of milk miRNAs on human gene expression. miRNA-containing milk exosomes affected the activity of reporter gene plasmids. When HEK-293 cells were cultured in media supplemented with milk exosomes providing 600 fmol/L miR-29b for 4 h, the activity of the miR-29b–dependent reporter

FIGURE 1 Plasma time curves of miR-29b (A), miR-1 (negative control), and miR-200c (B) after a milk meal in healthy adults. Values are means, n = 5. SEMs were omitted for clarity (compare with Table 1). miR, microRNA.
gene LUC-mir-29b decreased by 44 ± 13% (P = 0.04) compared with cells cultured in exosome-depleted culture media (Fig. 3A). Likewise, when HEK-293 cells were cultured in media supplemented with milk exosomes providing 1000 fmol/L miR-200c for 4 h, the activity of the miR-200c–dependent reporter gene LUC-mir-200c decreased by 17 ± 3% (P = 0.02) compared with cells cultured in exosome-depleted culture media.

Milk feeding caused changes in miRNA target gene expression in human PBMCs. The expression of RUNX2 was 31 ± 13% higher 6 h after the milk meal than at time 0 h (Fig. 3B; P = 0.04), consistent with the role of miR-29b as a positive regulator of RUNX2 (17). The difference in the expression of ZEB1 was not significantly lower 6 h after the milk meal than at time 0 h (Fig. 3B).

**miRNA depletion studies in mice.** Endogenous miRNA transcription does not compensate for dietary deficiency in C57BL/6 mice. When mice were fed an miRNA-defined diet for 4 wk, the miR-29b plasma concentration decreased by 61% from 152 ± 79 fmol/L in the miRNA-sufficient ExoPlus group to 60 ± 27 fmol/L in the miRNA-depleted ExoMinus group (P = 0.04). The expression of Kruppel-like factor 8 (KLF8) miRNA was 64 ± 32% lower in miRNA-depleted mice than in miRNA-sufficient mice (P = 0.04).

**Bioavailability of broccoli miRNAs in humans.** Broccoli sprouts contained 53 ± 16 and 42 ± 10 pmol/kg of miR-167a and miR-824, respectively. No change was observed in miR-167a plasma concentration in healthy adults after consumption of a broccoli sprout meal (13 ± 4 fmol/L at t = 0 h vs. 15 ± 7 fmol/L at t = 4 h; n = 4; P = 0.85). The concentration of miR-824 was below the detection limit (<1 fmol/L) at baseline or 4 h after broccoli sprout feeding in all of the 4 participants who were randomly chosen from our archived plasma samples (35).

**Discussion**

Some noncoding RNAs, including miRNAs, play essential roles in gene regulation in plant and animal kingdoms. To the best of our knowledge, this is the first study to report that humans absorb quantitatively meaningful amounts of miRNAs from nutritionally relevant amounts of cow milk. Our study provides unambiguous evidence that the amounts of miRNAs absorbed from milk are sufficient to alter human gene expression, i.e., miRNAs from 1 mammalian species can affect gene networks in another species. Our observation that endogenous miRNA synthesis cannot compensate for dietary deficiency is of particular interest, because it implies that a regular dietary miRNA intake may be important to prevent aberrant gene regulation.

Our observations are important from the perspective of maintaining human health on the basis of the following rationale. First, 245 miRNAs have been identified in cow milk (14). A preliminary analysis by sequence alignment suggests that the majority of the nucleotide sequences in bovine milk match the sequences of their human orthologs. We are in the process of developing algorithms for predicting human gene targets for bovine miRNAs, and initial assessments suggest that the number of target genes will exceed 11,000. Second, there is unambiguous evidence linking miRNAs with human health. For example, miR-29b promotes bone health through altering osteoblast and osteoclast differentiation (17,18), miR-200c decreases cancer risk by targeting the transcription factor ZEB1 (19,20), and miR-15b, miR-21, miR-27b, miR-34a, miR-106b, miR-130a, miR-155, miR-200c, and miR-223 have been implicated in immune function and Crohn disease (15,38). Third, supplementation and depletion of milk altered miRNA concentrations and gene expression in human PBMCs and in mouse liver. For example, the concentration of miR-200c was greater 6 h after a milk meal than at time zero in human PBMCs. Likewise, the expression of KLF8 miRNA was greater in livers from milk miRNA-sufficient mice than in those from milk miRNA-depleted mice. Fourth, milk is an important staple in many Western diets. For example, Americans consume large quantities of milk, despite a steady decline from ~237 pounds (108 kg) in 1987 to 195 pounds (88 kg) in 2012 (13,39). Note that a large

**FIGURE 2** Effects of milk microRNAs on the abundance of microRNAs in human peripheral blood mononuclear cells. Values are means ± SEMs, n = 5. *Different from hour 0, P < 0.05. miR, microRNA.
Bioavailability of milk microRNAs


